

FD'02

Course Syllabus for Biology 210-Biological Techniques  
Fall 2002  
Chaminade University of Honolulu

Meeting time & place: Monday 2:00-4:50 PM Henry Hall 13

Textbook: No text; several handouts including excerpts from the *Chemical Hygiene Plan at Chaminade University* and *Basic Laboratory Methods for Biotechnology* (Siedman & Moore). Additional materials on safety, protocols, techniques and instruments will be distributed during the semester.

Instructor: Dr. Joan Kuh

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*Office Hours:* WF 12-2, T 1-2 or by appointment

Course Description:

Biological techniques refers to the methodology, skills and instrumentation utilized in the natural sciences, specifically the *biological* sciences. This course was developed to provide experience and exposure to techniques and instrumentation for those students at the freshman and sophomore levels. Techniques include preparing solutions, pipetting, electrophoresis and manipulation of biological macromolecules. The aims of this course include: increasing student confidence and proficiency in biological laboratory skills that include safety considerations; preparation of students for skills necessary in future courses and/or careers; and preparation for assisting instructors at Chaminade University in laboratories and research projects. As a byproduct, you will be building *marketable* skills.

Course Objectives: The student is expected to demonstrate the following:

1. Ability to work safely in a biological laboratory. This requires 1) knowledge of the potential physical, chemical and biological hazards that can be encountered at CUH and in biological labs in general and 2) familiarity with the Chemical Hygiene Plan in place at CUH.
2. Working safely, cleanly and with consideration of other persons in the laboratory.
3. Proper operation and basic maintenance of instrumentation in a biological laboratory including autoclaves, balances, spectrophotometers, pipettes, centrifuges, balances, pH meters.
4. Basic techniques such as manipulation of microbes and macromolecules, pipetting, solution preparation, extracting protocols from publications, electrophoresis.
5. Participation in researching, designing and setting up a project with other individuals.
6. Proper and accurate laboratory procedure documentation as well as accurate analysis and reporting of data.

Grades will be based on the following criteria:

Assignments (9 total):	180 points
Safety Quiz	20 points
Class Project (reports* required)	50 points
Notebook (2 checks @ 25 points)	50 points
One practical midterm exam:	50 points
Final exam	100 points



*Tentative Schedule of Topics (all labs on Monday):*

Date	Topics	Assignments*
August 26*	Course Introduction Tour of Laboratories Safety Videos (HH 17)	I. Careers in Biology II. DAPI III. Basic Math Functions
September 9	Review Assignments Chemical Hygiene Plan (HH 17) Autoclaves/Distilling Water Glass and Plastic Labware	---Safety quiz next week! IV. Retrieving/analyzing MSDSs
September 16	Safety Quiz Balances/stir plates Documentation/Notebooks Preparing Solutions I Class Project discussion	V. Solutions
September 23	Preparing Solutions II Temperature/pH Filtration Measuring volume/pipetting Biohazards in the Lab (HH 17)	VI. pH/pipetting worksheet (due at end of lab)
September 30	Spectrophotometry Basic Statistics/Graphing Prepare plates/media for next week	VII. Statistics Problem Set
October 7*	Microbiological Techniques (plating, streaking, colony isolation) Making dilutions and titering	Practical Lab exam next meeting VIII. Titering bacterial cultures
October 21	Practical Laboratory Exam	
October 28	Macromolecules Centrifuges Plasmid DNA Isolation	
November 4	Gel Electrophoresis "Photodocumentation" Restriction Enzyme Analysis	IX. DNA quantitation/analysis
November 18	Wrap up Class Project	Class project report due next week.
November 25	Field Trip—UH Transgenics Lab UH Microscope Facility	
December 2	FINAL EXAM (accumulative) 2:00 – 4:50 PM	